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PATENT AND TRADEMARK OFFICE

**INFORMATION DISCLOSURE
STATEMENT**

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D. Saunders

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1643

Invention Title
**METHOD OF SCREENING FOR PROTEIN
INHIBITORS AND ACTIVATORS**

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Assistant Commissioner for Patents
Washington D.C. 20231

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SIR:

1. In accordance with the duty of disclosure under 37 C.F.R. § 1.56 and in conformance with the procedures of 37 C.F.R. §§ 1.97(c) and 1.98 and M.P.E.P. § 609, Applicant hereby brings the references listed on the attached modified PTO Form No. 1449 to the attention of the Examiner. It is respectfully requested that the references be expressly considered during the prosecution of this application, and that the references be made of record therein and appear among the "References Cited" on any patent to issue therefrom.
2. Consistent with the policy that we have followed throughout the prosecution, we bring to the attention of the Examiner the following reference which was recently brought to our attention:
(1) Balzarini, J. et al. (1985) FEBS Lett. 185:95-100.
3. No item of information contained in this information disclosure statement (IDS) was cited in a communication from a foreign patent office in a counterpart foreign application, and to the knowledge of the undersigned after making reasonably inquiry, no item of information contained in this IDS was known to any individual designated in § 1.56(c) for than three months prior to filing of this IDS.
4. It is believed that no fees are due in connection with this Information Disclosure Statement. However, should any fees be due, the Commissioner is authorized to charge Deposit Account No. 11-0600 for such fees. A duplicate copy of this communication is enclosed for charging purposes.

Dated: April 24, 2001

By:


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Murine mammary FM3A carcinoma cells transformed with the herpes simplex virus type 1 thymidine kinase gene are highly sensitive to the growth-inhibitory properties of (E)-5-(2-bromovinyl)-2'-deoxyuridine and related compounds

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Murine mammary carcinoma (FM3A TK⁻/HSV-1 TK⁺) cells, which are thymidine kinase (TK)-deficient but have been transformed with the herpes simplex virus type 1 (HSV-1) TK gene are inhibited in their growth by (E)-5-(2-bromovinyl)-2'-deoxyuridine (BVDU), (E)-5-(2-iodovinyl)-2'-deoxyuridine (IVDU) and (E)-5-(2-bromovinyl)-2'-deoxycytidine (BVDC) at 0.5, 0.5 and 0.8 ng/ml, respectively; i.e., a concentration 5000 to 20000-fold lower than that required to inhibit the growth of the corresponding wild-type FM3A/0 cells. Hence, transformation of tumor cells with the HSV-1 TK gene makes them particularly sensitive to the cytostatic action of BVDU and related compounds.

<i>Murine mammary FM3A carcinoma</i>	<i>Thymidine kinase gene</i>	<i>Herpes simplex virus type 1</i>	<i>Cytostatic activity</i>
	(E)-5-(2-Bromovinyl)-2'-deoxyuridine		

INTRODUCTION

Recently, several potent and selective antiviral agents have been developed, i.e. 9-(2-hydroxyethoxymethyl)guanine (acyclovir, ACV) [1], 1-[(1,3-dihydroxy-2-propoxy)methyl]guanine (DHPG) [2,3], (E)-5-(2-bromovinyl)-2'-deoxyuridine (BVDU) [4] and its closely related analogues (E)-5-(2-iodovinyl)-2'-deoxyuridine (IVDU) [4], (E)-5-(2-bromovinyl)-2'-deoxycytidine (BVDC) [5] and (E)-5-(2-bromovinyl)-1-β-D-arabinofuranosyluracil (BVaraU) [6,7], 1-(2'-fluoro-2'-deoxy-1-β-D-arabinofuranosyl)-5-iodocytosine (FIAC) [8,9], 1-(2'-fluoro-2'-deoxy-1-β-D-arabinofuranosyl)-5-methyluracil (FMAU) [9,10] and phosphonoformic acid (PFA) [11]. The selectivity of these com-

pounds as inhibitors of herpes simplex virus (HSV) primarily depends upon a specific activation (phosphorylation) by the HSV-encoded thymidine (dThd) kinase (TK), except for PFA [12]. Indeed, upon HSV infection, a virus-specified TK is induced, which differs from human cytosol and mitochondrial TK in its physical, immunological and kinetic behavior [13]. It is endowed with deoxycytidine (dCyd) kinase activity and shows a much greater affinity for several nucleoside analogues including ACV [14], DHPG [15], BVDU [16], IVDU [16], BVDC [16], BVaraU [17], FIAC [17] and FMAU [17], than the cellular (cytosol) TK. The fact that these compounds are intensively phosphorylated by the viral TK, and only to a very limited, and often undetectable, extent by the cellular TK, apparently accounts for their low level of toxicity for the uninfected host cell.

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We have previously described a thymidylate synthetase (TS)-deficient mutant cell line (designated FM3A/TS⁻) which was derived from mutagenized murine mammary carcinoma FM3A/0 cells [18-21]. This mutant cell line, which is auxotrophic for dThd appeared very useful to distinguish those pyrimidine nucleoside analogues that are, like dThd, incorporated into host cell DNA and stimulatory for cell growth from those nucleoside analogues that are not [20,21]. We have now constructed a TK-deficient FM3A cell line (FM3A TK⁻) which was subsequently transformed by a fragment of HSV-1 DNA containing the gene for TK (hence designated FM3A TK⁻/HSV-1 TK⁺). A broad variety of both selective and non-selective antiherpetic drugs were evaluated for their inhibitory effects on the growth of FM3A/0 and FM3A TK⁻/HSV-1 TK⁺ cells. Those compounds whose anti-herpes activity does not depend on a specific phosphorylation by the HSV-1 TK proved equally inhibitory to the proliferation of FM3A/0 and FM3A TK⁻/HSV-1 TK⁺ cells. Some (but not all) of the compounds, whose inhibitory effects on HSV-1 replication depend on a specific phosphorylation by the viral TK were much more inhibitory to the growth of FM3A TK⁻/HSV-1 TK⁺ than FM3A/0 cells. For example, BVDU inhibited the growth of FM3A TK⁻/HSV-1 TK⁺ and FM3A/0 cells at 0.5 ng/ml and 11.4 µg/ml, respectively. Thus, transformation of the FM3A cells with the HSV-1 TK gene increased the cytostatic activity of BVDU 22800-fold.

2. MATERIALS AND METHODS

2.1. Cells: growth conditions, origin and selection

Murine FM3A cells (subclone F28-7), originally established from a spontaneous mammary carcinoma in a C3H/He mouse [22] and designated FM3A/0, were grown in 75-cm² tissue culture flasks (Sterilin, Teddington, England) in Eagle's MEM, supplemented with 10% (v/v) inactivated fetal calf serum (Gibco Bio-Cult, Glasgow, Scotland), 2 mM L-glutamine (Flow Laboratories, Irvine, Scotland) and 0.075% (w/v) NaHCO₃ (Flow Laboratories). FM3A/TS⁻ cells were maintained in the same culture medium, supplemented with 20 µM dThd [20,21].

The FM3A TK⁻/HSV-1 TK⁺ cell line, which lacks host cell TK activity but contains the HSV-1

TK gene, was originally derived from a subclone of FM3A/0 cells, made deficient for host cell TK by selection in the presence of 5-bromo-2'-deoxyuridine (BDU). The HSV-1 TK gene was introduced by DNA-mediated gene transfer using pBR322 plasmid containing the 3.6-kb *Bam*HI fragment of HSV-1 DNA at a *Bam*HI site. The plasmid clone used for transformation did not contain the genes for viral DNA polymerase or ribonucleotide reductase.

From the FM3A TK⁻/HSV-1 TK⁺ cells a TS-deficient subclone was derived as in [22]. Shortly, FM3A TK⁻/HSV-1 TK⁺ cells were mutagenized with 1 µg/ml *N*-methyl-*N'*-nitro-*N'*-nitrosoguanidine for 3 h at 37°C. The cells were grown for 5 days in the presence of 10⁻⁵ M dThd, and then plated on agarose in the presence of 10⁻⁶ M 5-methyltetrahydrofolate, 2 × 10⁻⁷ M methotrexate and 10⁻⁵ M dThd. FM3A TK⁻/HSV-1 TK⁻/TS⁻ colonies were isolated after 10 days. FM3A TK⁻/HSV-1 TK⁺ and FM3A TK⁻/HSV-1 TK⁻/TS⁻ cells were cultured in the same medium as FM3A/0 and FM3A/TS⁻ cells, i.e., in the absence or presence of 20 µM dThd, respectively.

2.2. Test compounds

The source of the test compounds was as follows: BVDU and IVDU, synthesized by R. Busson and H. Vanderhaeghe of the Rega Institute for Medical Research (Katholieke Universiteit Leuven, B-3000 Leuven), following a modification of the method described by Jones et al. [23]; BVaraU, provided by H. Machida (Yamasa Shoyu Co., Choshi, Japan), see also [24]; BVDC, provided by R.T. Walker (University of Birmingham, Birmingham, England); see also [23]; ACV, Burroughs Wellcome Co. (Research Triangle Park, NC); the 8-ido-derivative of ACV (IACV), provided by M.J. Robins (University of Alberta, Edmonton, Canada), see also [25]; FIAC and FMAU, provided by J.J. Fox (Sloan-Kettering Institute, New York, NY), see also [9,10]; DHPG, provided by J.P.H. Verheyden (Syntex Research, Palo Alto, CA); 5-(2-chloroethyl)-2'-deoxyuridine (CEDU), provided by B. Rosenwirth (Sandoz Forschungsinstitut, Vienna, Austria); see also [26]; 1-β-D-arabinofuranosylthymine (araT), provided by H. Machida (Yamasa Shoyu Co., Choshi, Japan); 5-fluoro-2'-deoxyuridine (FDU), Aldrich (Milwaukee, WI); BDU, Sigma Chemical Com-

pany (St. Louis, MO); 5-iodo-2'-deoxyuridine (IDU), Sigma; 5-trifluoromethyl-2'-deoxyuridine (TFT), P-L Biochemicals (Milwaukee, WI); 5-ethyl-2'-deoxyuridine (EDU), Robugen GmbH (Esslingen, FRG), see also [27,28]; 5-propyl-2'-deoxyuridine (PDU), [29]; 9- β -D-arabinofuranosyladenine (araA), Sigma; 1- β -D-arabinofuranosylcytosine (araC), Upjohn Company (Puurs, Belgium); and PFA, provided by B. Öberg (Astra Läkemedel AB, Södertälje, Sweden). The therapeutic potentials of these anti-herpes agents have been reviewed in [30,31].

2.3. Inhibition of tumor cell growth

All assays were performed in 96-multiwell microtest plates (Falcon, Becton Dickinson, Oxnard, CA). To each well were added 5×10^4 cells and varying amounts of the test compounds. The cells were then allowed to proliferate at 37°C in a humidified, CO₂-controlled atmosphere. In preliminary assays it was assessed that the growth of the cells was linear for up to 72 h. In the cell growth-inhibition experiments, the incubation was stopped at 48 h. The cells were then enumerated in a Coulter Counter (Coulter Electronics, Harpenden, England). The cell growth-inhibitory effects of the test compounds are expressed in ID_{50} , or the inhibitory dose required to reduce the final cell number by 50%.

3. RESULTS AND DISCUSSION

3.1. Inhibitory effects of anti-herpes agents on the proliferation of FM3A/0 and FM3A TK⁻/HSV-1 TK⁺ cells

A wide series of anti-herpes agents were evaluated for their inhibitory effects on the growth of FM3A/0 and FM3A TK⁻/HSV-1 TK⁺ cells (table 1). Based on whether the compounds act as preferential substrates for the HSV-1 TK [12], distinction was made between selective and non-selective anti-herpesvirus agents (although it should be recognized that some of the compounds belonging to the latter category, i.e., araA and PFA, may interact specifically with the viral DNA polymerase).

Among those compounds whose anti-herpes activity depends on a specific phosphorylation by the viral TK, several congeners were much more inhibitory to the proliferation of FM3A TK⁻/HSV-1

TK⁺ than of FM3A/0 cells; i.e., BVDU (22800-fold), BVDC (20250-fold), IVDU (5600-fold), araT (678-fold) and DHPG (130-fold). Clearly, phosphorylation of these compounds by the viral TK is required for their inhibitory effect on the growth of HSV-1 transformed cells. Although obligatory, this phosphorylation may not be sufficient for a cytostatic effect of the anti-herpes agents on HSV-1 transformed cells, since several other compounds, ACV, FMAU, EDU and BVaraU, which also

Table 1

Inhibitory effects of anti-herpes agents on the proliferation of FM3A/0 and FM3A TK⁻/HSV-1 TK⁺ cells

Compound ^a	ID_{50} ($\mu\text{g/ml}$)		ID_{50} (FM3A/0)
	FM3A/0	FM3A TK ⁻ / HSV-1 TK ⁺	
Selective anti-herpes agents			
BVDU	11.4	0.0005	22800
BVDC	16.2	0.0008	20250
IVDU	2.80	0.0005	5600
araT	238	0.351	678
DHPG	37.5	0.288	130
FMAU	5.92	1.20	4.9
ACV	27.2	7.32	3.7
IACV	180	74	2.4
FIAC	>100	>10 ^b	—
CEDU	35.4	>10 ^b	—
EDU	9.47	>10 ^b	—
BVaraU	>300	>300	—
PDU	>1000	>1000	—
Non-selective anti-herpes agents			
araA	15.4	3.72	4.1
TFT	0.007	0.004	1.7
PFA	221	137	1.6
araC	0.211	0.150	1.4
FDU	0.0005	0.0006	0.8
BDU	38.5	620	0.06
IDU	7.38	454	0.02

^a Considered as 'selective' if preferentially phosphorylated by the HSV-1 TK [12]

^b ID_{50} value could not be measured accurately within the concentration range of 10–1000 $\mu\text{g/ml}$